N-(2-Pyridyl)glycine hydrochloride: mp 200-205 °C dec (lit.¹⁹ mp 190 °C).

N-(2-Pyridyl)glycine: mp 170–174 °C (lit.¹⁹ mp 175 °C). **N**-(2-Pyridyl)-α-alanine hydrochloride: mp 198–210 °C dec; ¹H-NMR (DMSO- d_6 , 60 MHz) δ 1.5 (d, 3 H, J = 7.8 Hz, Me), 4.8 (q, 1 H, J = 7.8 Hz, CH), 7.4 (m, 2 H), 7.7–8.1 (m, 2 H), 8.8–9.6 (m, 1 H); IR (KBr) ν 1730, 1650 (CO) cm⁻¹. Anal. Calcd for C₈H₁₁N₂O₂Cl: C, 47.40; H, 5.43; N, 13.83, Cl, 17.53. Found: C, 47.12; H, 5.67; N, 13.96; Cl, 17.79.

N-(2-Pyrimidinyl)-α-alanine: mp 180–183 °C (H₂O); ¹H NMR (DMSO- d_6 , 60 MHz) δ 1.3 (d, 3 H, J = 7.0 Hz, Me), 4.2 (qt, 1 H, $J_1 = J_2 = 7.0$ Hz, CH), 6.3 (t, 1 H), 7.0 (d, 1 H, NH), 7.9 (d, 2 H); IR (KBr) ν 1695 (CO) cm⁻¹. Anal. Calcd for C₇H₉N₃O₂: C, 50.30; H, 5.38; N, 25.10. Found: C, 49.98; H, 5.65; N, 25.04. The corresponding hydrochloride salt was a viscous yellow oil.

N-(2-Thiazolyl)-α-alanine hydrochloride: mp 170–190 °C dec; ¹H NMR (DMSO- d_6 , 60 MHz) δ 1.5 (d, 3 H, J = 7.2 Hz, Me), 4.7 (q, 1 H, J = 7.2 Hz, CH), 7.0 (d, 1 H), 7.3 (d, 1 H); IR (KBr) ν 1740 (CO) cm⁻¹. Anal. Calcd for C₆H₉N₂O₂SCl: C, 34.53; H, 4.31; N, 13.43; S, 15.34; Cl, 17.02. Found: C, 34.34; H, 4.36; N, 13.58; S, 15.32; Cl, 17.29.

N-(2-Pyrimidinyl)phenylglycine hydrochloride: mp 185–190 °C; ¹H NMR (DMSO- d_6 , 60 MHz) δ 5.7 (s, 1 H, CH), 7.0 (t, 1 H), 7.2–8.0 (m, 5 H), 8.6 (d, 2 H); IR (KBr) ν 1730, 1640 cm⁻¹. Anal. Calcd for C₁₂H₁₂N₃O₂Cl: C, 54.23; H, 4.52; N, 15.82; Cl, 13.37. Found: C, 54.51; H, 4.43; N, 15.52; Cl, 13.12.

N-(2-Thiazolyl)phenylglycine hydrochloride: mp 86–92 °C; ¹H NMR (DMSO- d_6 , 60 MHz) δ 5.8 (s, 1 H, CH), 6.9 (d, 1

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H), 7.1–7.7 (m, 6 H); IR (KBr) ν 1730 (CO) cm⁻¹. Anal. Calcd for C₁₁H₁₁N₂O₂SCl: C, 48.79; H, 4.06; N, 10.35; S, 11.83; Cl, 13.12. Found: C, 48.52; H, 4.37; N, 10.32; S, 11.90; Cl, 12.87.

Hydrolysis of α -Amino Ester 4g. Following the general procedure for the hydrolysis of α -amino esters 4, 0.2 g of compound 4g in 5 N hydrochloric acid (2 mL) were heated under reflux for 3 h. After this time the solvent was removed under vacuo and the white crystalline solid thus obtained was treated with saturated sodium carbonate solution (4 mL). In this way 0.19 g of a yellow powdered solid was obtained. This compound was identical with an authentic sample of 2-phenyl-1*H*-imidazo[1,2-*a*]pyridinium-3-olate (2, X = CH).¹³

Acknowledgment. Support for this research under Grant PB87-0064-C03-00 from the DGICYT (M.E.C., Spain) is gratefully acknowledged. M.A.S. thanks the Ministerio de Educación y Ciencia (Spain) for a F.P.I. fellowship.

Registry No. 2 X = CH, Ar = Ph, 106492-20-0; 3 R = Me-HClO₄, 126190-30-5; 4a, 100377-28-4; 4b, 53051-79-9; 4c, 126190-19-0; 4d, 126190-20-3; 4e, 28036-34-2; 4f, 126190-21-4; 4g, 126190-22-5; 4h, 126190-23-6; 4i, 126190-24-7; 4j, 126190-25-8; 4k, 126190-26-9; 4l, 126190-27-0; 4m, 126190-28-1; 4n, 126190-29-2; 5a, 52946-88-0; 5a-HCl, 112656-88-9; 5d-HCl, 122886-09-3; 5k, 126190-31-6; 5l-HCl, 126190-33-8; 5m-HCl, 126190-32-7; 5n-HCl, 126190-34-9; glyoxal, 107-22-2; pyruvaldehyde, 78-98-8; phenylglyoxal, 1074-12-0; 2-aminopyridine, 504-29-0; 4-methyl-2aminopyridine, 695-34-1; 5-chloro-2-aminopyridine, 1072-98-6; 2-aminopyrimidine, 109-12-6; 2-aminothiazole, 96-50-4; 2aminopyrazine, 5049-61-6.

Synthesis of Complex 6'-Alkynyl-6'-dethia Nucleoside Analogues of S-Adenosylhomocysteine as Potential Inhibitors of Methyltransferases

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Received November 20, 1989

The synthesis of complex acetylenic nucleosides containing a 5',6' carbon-carbon bond has been investigated. Two synthetic routes were studied, both starting with an α -substituted butyrolactone. The lactone could be converted to an β -keto sulfone and then to the corresponding enol phosphate diphenyl ester. Reductive elimination to yield the desired aryl acetylene could not be effected without some overreduction to the substituted olefin. Alternatively, the lactone could be converted to an appropriately protected γ -hydroxy aldehyde which was then homologated to a terminal acetylene. The successful synthesis of a prototypic member of the target class of compounds involved a Pd-mediated coupling of an aryl iodide to the appropriately functionalized terminal acetylene. The resulting aryl acetylene was then further elaborated to the target amino acid derivative; viz, a 6'-alkynyl-6'-dethia analogue of S-adenosylhomocysteine.

Introduction

The reaction of a variety of cellular nucleophiles with electrophilic biochemical alkylating agents such as Sadenosylmethionine (AdoMet, 1) or its decarboxylated derivtive, dcAdoMet, 2, is extremely important in cellular metabolism. AdoMet acts as a methyl donor in the methylation of molecules as diverse as catecholamines and nucleic acids.¹ Similarly dcAdoMet acts as an aminopropyl donor in the biosynthesis of the cationic polyamines, spermidine and spermine, which are known to interact with anionic nucleic acids² and otherwise affect cell growth and differentiation.



Stereochemical studies of the reactions catalyzed by methyltransferases, e.g., catechol O-methyltransferase (COMT),³ and aminopropyltransferases, e.g., spermidine

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synthase,⁴ have led to the conclusion that these alkyltransfer reactions involve a direct $S_N 2$ attack of the nucleophilic substrate on the electrophilic substrate. It is not yet clear whether general base catalysis of intramolecular alkyl-transfer reactions, observed in several model reactions,⁵ is operative in the case of the enzyme-catalyzed reactions. Kinetic studies have shown that both COMT and spermidine synthase are severely inhibited by accumulation of the thioether products, S-adenosylhomocysteine (AdoHcy) and 5'-deoxy-5'-(methylthio)adenosine, respectively.⁶ A large number of laboratories have been involved in the synthesis and biological evaluation of a wide variety of structural analogues of these naturally occurring product inhibitors.⁶ Although many of these compounds have been of considerable utility in the study of intracellular alkyl-transfer reactions, it is unlikely that one can obtain specific inhibition of a particular alkyltransfer reaction of interest by using a structural analogue of a product which is common to all such reactions. As a result, we have undertaken the synthesis of a series of "multisubstrate adduct" inhibitors of methyltransferases7 and aminopropyltransferases.^{8,9} In the case of the aminopropyltransferases, the multisubstrate adduct inhibitors have proven to be extremely potent inhibitors and ex-

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quisitely specific for the target enzyme, spermidine synthase¹⁰ or spermine synthase.¹¹ These compounds have been used extensively in the study of the regulation of polyamine biosynthesis in vitro.¹²

There is considerable interest in the development of selective COMT inhibitors as adjuncts to the dopamine precursor, L-DOPA, in the treatment of Parkinson's disease.13 Unfortunately, our initial attempts to design synthetic multisubstrate adduct inhibitors for a methyltransferase such as COMT have not met with as much success.⁷ Encouraged by the results with aminopropyltransferases briefly summarized above, we have undertaken the synthesis of a new class of compounds designed to be specific inhibitors of individual methyltransferases of interest. Considering the mechanistic data obtained by ourselves (loc. cit.) and others,¹⁴ we have proposed a transition state 3 for the COMT-catalyzed reaction in which the 3-hydroxyl group attacks the electrophilic methyl group in a classic $S_N 2$ mechanism. A reasonable mimic of this array is represented by 5 or 6, incorporating surrogates for the COMT nucleophilic substrates, 3,4-dihydroxybenzoic acid or epinephrine, respectively, and the electrophilic methyl donor, AdoMet, connected via an acetylenic moiety. In this paper, we describe the synthesis of 4, a prototypic member of this new class of compounds, viz, 6'-alkynyl-6'-dethia analogues of AdoHcy. In principle, it should be possible, using the synthetic methods described in this paper, to place a variety of substituents on

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^aReagents: (a) $TrO(CH_2)_3P^+Ph_3Br^-$ (8), *n*-BuLi, THF; (b) H_2/PtO_2 , EtOH, 60 psi; (c) Li/NH_3 ; (d) NH_4OH , aqueous MeOH; (e) $(PhO)_3P^+CH_3L^-$, CH_2Cl_2 ; (f) PhCH— NCH_2CO_2Me/LDA , THF/HMPA; (g) 80% HCO_2H ; (h) K_2CO_3 , MeOH/H₂O.

the alkyne which mimic the nucleophilic component of the ternary complex for any one of the multitude of Ado-Met-dependent specific methyltransferases.



Results and Discussion

Synthetic Strategy (Scheme I). The two routes which were investigated to develop a general synthesis of 4-6 are outlined retrosynthetically in Scheme I. The synthesis of the target compounds via route A involves the reductive elimination of a phosphoenol arylsulfonyl nucleoside. This nucleoside could be derived from the corresponding β -ketosulfonyl nucleoside, and this β -ketosulfonyl nucleoside could be synthesized by the coupling of an aryl sulfonyldianion to a lactone nucleoside or its equivalents. Via route B, 4-6 could be obtained by the arylation of a terminal acetylenic nucleoside with an aryl iodide. The terminal acetylenic nucleoside could be derived from the corresponding aldehyde nucleoside, and the aldehyde nucleoside could be synthesized from the same lactone nucleoside. It was envisioned that the amino acid portion of the target compounds could be elaborated by

addition of an amino acid equivalent to an electrophilic carbon, e.g., alkyl halide or sulfonate on the side chain. Therefore, the synthesis of the 6'-dethia analogue of AdoHcy (15) was investigated as a model for this type of synthetic elaboration. In addition to the synthetic purpose, 6'-dethia AdoHcy is of use as a less complex compound, lacking the catecholamine portion of 6, in the enzyme inhibition studies.

Synthesis of the 6'-Dethia Analogue of AdoHcy (Scheme II). The synthesis of the 6'-dethia analogue of AdoHcy 15 is shown in Scheme II. The phosphonium bromide 8 used in the first step was prepared by tritylation of 3-hydroxypropyl bromide with trityl bromide, DMAP, and Et₃N in DMF,¹⁵ followed by the triphenylphosphine treatment of the resulting O-tritylated intermediate in refluxing benzene. Condensation of 8 with the anhydrous 5'-aldehyde nucleoside 7^{16} in the presence of *n*-BuLi in THF from -78 °C to 0 °C smoothly produced the bondextended cis and trans olefinic nucleoside 9 (about 2:1 ratio of cis:trans) in 89–95% yield. Detritylation and reduction of the double bond of 9 by catalytic hydrogenation was not straightforward. Several attempts to effect these two reactions either simultaneously or stepwise using catalytic hydrogenation and/or treatment with Li/NH₃ or Na/NH₃ failed to give the desired alcohol, 11, in acceptable yields $^{17-19}$ Hydrogenation of 9 with Adams catalyst (PtO₂) at 60 psi of hydrogen pressure for 3 days afforded only a 24% yield of the desired N^6 -benzoyl hydroxypropyl nucleoside 10a along with 68% of the N^6 -benzoyl O-tritylprotected nucleoside 10b and 7% of the debenzoylated O-trityl-protected nucleoside 10c. It is known that palladium hydroxide on carbon²⁰ is an effective catalyst for

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^aReagents: (a) SOCl₂; (b) 5-R₂-4-R₃-2-R₄-C₆H₂-CH₂SO₂Tol(16 or 17)/BuLi, THF; (c) (MeO)₂CHN(Me)₂, DMF; (d) ClP(O)(OPh)₂, DMAP, Et_3N , CH_3CN ; (e) HOAc/ $EtOH/H_2O$ (v/v, 3/2/1); (f) 5.65% Na-Hg, THF/DMSO.

debenzylation, and Hanessian et al.²¹ have used this catalyst to achieve a successful detritylation in their studies. Using this catalyst (32% by weight) for the detritylation of the mixture of 10b (major) and 10c (minor) in EtOH under 50 psi of hydrogen pressure for 3 days gave 31% of 10a and 24% of 11. Debenzoylation of the isolated 10a with ammonium hydroxide in methanol²² at room temperature for 18 h produced 11 in 89% yield. Ultimately, the conditions described in the Experimental Section were found to be optimal for the conversion of 9 to 11 without purification of the intermediates 10a-c. The hydroxypropyl nucleoside 11 was converted to the iodopropyl nucleoside 12 in 75-77% yield by reacting 11 with methyltriphenoxyphosphonium iodide in methylene chloride.²³ Treatment of the benzylidene-derived methyl glycinate²⁴ with 1 equiv of lithium diisopropylamide (LDA) followed by 1 equiv of 12 in THF and HMPA from -78 °C to room temperature provided the desired adduct 13 in 95% crude yield. ¹H NMR and TLC studies of the crude 13 indicated that the conversion of 12 to 13 was completed. Purification of 13 was not attempted due to its instability toward silica gel chromatography. Acid hydrolysis of the crude 13 with 90% trifluoroacetic acid at room temperature for 20 min²² in order to cleave both the isopropylidene and the ben-

zylidene groups also caused a great deal of depurination. Better results were achieved by using 80% formic acid²⁵ at room temperature for 5-6 h which gave a much cleaner formate salt of 14. Pure 14 (free of the formate salt) could be obtained by the purification of crude 14 on preparative reverse-phase (C-18) HPLC. However, it is more convenient to convert the crude 14 directly to the final product 15. Thus, base hydrolysis of the methyl ester group of 14 with 5 equiv of K_2CO_3 in aqueous MeOH²⁵ at room temperature overnight produced the crude 6'-dethia AdoHcy, 15. A portion of crude 15 was purified on preparative reverse-phase (C-18) HPLC to give pure 15 in 22% overall vield from 12.

These results, especially those involving the conversion of the hydroxypropyl nucleoside 11 to the 6'-dethia analogue of AdoHcy 15, and the experience obtained from the preparative reverse-phase HPLC separations of polar molecules provided an experimental basis for the synthesis and purification of the prototype 4 in the route B approach described below.

Route A Approach (Scheme III). The methodology used in this approach for the construction of the carboncarbon triple bond was developed independently by Bartlett's²⁶ and Lythgoe's²⁷ groups in 1978. They reported

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the synthesis of acetylenes in good yields from carboxylic acid derivatives (esters, acid chlorides, and lactone) and alkyl phenyl sulfones. However, similar chemistry with benzyl phenyl sulfones was not explored. For the synthesis of the proposed inhibitors 5 and 6 via this approach, one needs a disubstituted benzylphenylsulfone to couple with a lactone nucleoside. In order to investigate the feasibility of using a benzyl phenyl sulfone in this approach, several model reactions were first carried out (eq 1). When re-



action of α -methyl- γ -butyrolactone, 18a, with benzyl phenyl sulfone gave only poor yields of the desired β -keto sulfone, even under forcing conditions,²⁸ the more electron-rich *p*-methoxybenzyl phenyl sulfone, 16, was investigated. More satisfactory yields were obtained in the reaction of 16 and 18a, leading to an equilibrium mixture of the desired β -keto sulfone 19a and the hemiketal 20a. Unfortunately, when the lactone nucleoside 18b²⁹ was allowed to react with the α,α -dilithio derivative of 16 in THF (-78 °C to room temperature) only about 3% of the desired β -ketosulfonyl nucleoside 19b was isolated.

In order to investigate the reactions of better electrophiles, the acid chlorides 22a and 22b were prepared from the corresponding carboxylic acids 21a³⁰ and 21b,²⁹ respectively, by thionyl chloride treatment (see Scheme III). Condensation of the acid chloride 22a with the dianion of 16 in THF (-78 °C to -20 °C) gave the β -keto sulfone 23a in up to 67% yield. In a similar manner, the β -ketosulfonyl nucleoside 23b was obtained in up to 76% yield from 22b and 16. Condensation of 22a with the dianion of the 2methoxy-5-methylbenzyl tolyl sulfone 17 furnished the β -keto sulfone 23c in up to 56% yield. However, when 22b was reacted with the dianion of 17, the yields for the desired 23d ranged only from 19 to 52%. In most of the cases, the yields were in the range of only 30%. Reaction of the dianion derived from 17 with 2-ethylhexanoyl chloride gave the corresponding β -keto sulfone in 80% yield, thus establishing the efficacy of this anion in the synthesis of less complex β -keto sulfones. The *p*-methoxybenzyl keto sulfones, 23a and 23b, were further converted to their corresponding enol phosphates. Thus, enol phosphorylation of 23a with diphenyl phosphorochloridate, 4-(dimethylamino)pyridine (DMAP), and triethylamine in CH₃CN at 70 °C for 30 min produced the enol phosphate 24a in 78% yield. Reductive elimination of 24a with 5.65% Na-Hg in THF and DMSO furnished the disubstituted alkyne 25a in 68% yield. The alkyne 25a so obtained was contaminated with a trace amount (<5%) of the overreduced olefin 26a, which was inseparable from 25a.

In order to convert 23b to 24b, one first had to protect the N^6 -amino group of the adenosyl moiety of 23b.³¹ Treatment of 23b with N.N-dimethylformamide dimethyl acetal in DMF³² produced the N^6 -imino-protected β -ketosulfonyl nucleoside 23e. Due to the instability of 23e toward silica gel chromatography, it was used immediately without purification in the next step. Enol phosphorylation of the crude 23e with diphenyl phosphorochloridate afforded the N^6 -imino-protected enol phosphate nucleoside 24e. Again without purification, 24e was deprotected at its N^6 -imino group selectively with acetic acid in aqueous ethanol to achieve the desired enol phosphate nucleoside 24b in 38% overall yield from 23b. Unfortunately, reductive elimination of 24b (0.1 mmol) with 5.6% Na-Hg in THF and DMSO produced a 42% yield of an unseparated mixture of the desired alkyne nucleoside 25b and the overreduced olefin nucleoside 26b. The ratio of 25b to 26b could not be determined using ${}^{1}H$ NMR because of the complexity of the spectrum.³³ Our investigation of the route A approach has revealed several drawbacks as described above. However, this route is effective in affording nonnucleoside aryl acetylenes such as 25a in good yield (41% overall from 21a).

Route B Approach (Scheme IV). One of the two key steps in the route B approach is the conversion of an aldehyde to a terminal acetylene. Corey and Fuchs reported in 1972^{34} the synthesis of terminal acetylenes from the corresponding aldehydes via dibromo olefins. This method was adopted by Jones et al.³⁵ in the synthesis of the chain-extended acetylenic nucleoside from the corresponding N⁶-benzoyl-2',3'-isopropylidene-adenosine 5'aldehyde. Based on the compatibility of a nucleoside with the Corey and Fuchs chemistry, as demonstrated by Jones et al., this method was investigated. Reduction of the lactone 18a with 1.2 equiv of DIBAL in toluene and THF gave the lactol 29a (eq 2). Without isolation, 29a was



immediately treated with TBDPSiCl to afford the desired O-silyl-protected aldehyde **30a** in 26% overall yield from **18a**. The undesired **31a** was not observed. Similarly, reduction of **18b** with 2.5-3.5 equiv of DIBAL in toluene

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 ⁽³⁰⁾ The acid 21a was prepared in three steps from 18a in a similar manner to that described²⁹ for the synthesis of 21b from 18b.

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⁽³³⁾ The mass spectrum of the mixture gave a ratio of 45:55 for the molecular ions of **25b** (717 amu) vs **26b** (719 amu).

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and THF smoothly afforded the lactol nucleoside 29b in 62-86% yield. Silvlation of 29b with TBDPSiCl under various conditions frequently gave an inseparable mixture of the desired O-silvlated aldehyde nucleoside 30b and the undesired O-silylated lactol nucleoside 31b. Among the many reaction conditions studied,³⁶ the best result obtained was 88% total yield with a ratio of 30b:31b equal to 73:27. A similar problem was also observed by Nicolaou and Magolda³⁷ in their silvlation of a six-membered ring lactol.

Treatment of the mixture of 30b and 31b with CBr_4 (2 equiv), PPh₃ (2 equiv), and Zn dust (2 equiv) in CH_2Cl_2 resulted in quantitiative recovery of the unreacted starting mixture. The consumption of the in situ generated phosphorus ylide, $Ph_3P=CBr_2$, by the deprotonation of N^6 -amide proton of the purine base might be the cause of this failure. Therefore, treatment of the same mixture of 30b and 31b with 6 equiv of CBr_4 , 6 equiv of PPh_3 , and 6 equiv of Zn in CH₂Cl₂ smoothly gave the desired dibromo olefinic nucleoside 32b in 66% yield, corrected for the recovery of the inert silvlated lactol 31b. The conversion of 32b to the corresponding terminal acetylenic nucleoside, **33b** (eq 3), was first attempted with 4 equiv of *n*-butyl-



lithium (2 of the 4 equiv of n-BuLi were used to prevent the interference arising from the deprotonation of the N^6 -amide and possibly the C-8³⁸ protons of the adenine ring) in THF (-78 °C to 0 °C). These reaction conditions produced a very complex crude mixture as indicated by its TLC analysis. After extensive efforts to effect a chromatographic separation of this mixture, less than 22% vield of the debenzoylated terminal acetylenic nucleoside 33c and a 3% yield of the α -isomer of the N⁶-benzovlated terminal acetylenic nucleoside were isolated.³⁹ Corey and Fuchs also used 1.5% lithium amalgam in THF as an alternative method to convert those dibromo olefins which are sensitive to alkyllithium to the desired terminal acetylenes. However, numerous attempts to employ 1.5% Li-Hg in the conversion of 32b to 33b gave only a complex mixture of products.³⁶ In conclusion, the results obtained from both methods (i.e. n-BuLi and 1.5% Li-Hg), indicate that routine synthesis of the terminal acetylenic nucleoside is not practical using the method of Corey and Fuchs.

Better results were obtained when the dimethyl (diazomethyl)phosphonate 3440 was used for the one-step conversion of 30b to 33b. Treatment of the mixture of 30b and 31b with 34 in the presence of potassium tert-butoxide in THF from -78 °C to room temperature smoothly produced the desired N⁶-benzovlated terminal acetylenic nucleoside 33b in 56-77% yield, corrected for the recovery of the inert 31b (Scheme IV). The characteristic ¹H NMR identification of 33b is the unique two sets of doublets due to the acetylenic CH (ca. 2:1 ratio of diastereomers) at ca. 2.05 ppm with an allylic coupling constant of 2.4 Hz. The arylation of terminal acetylenes with aryl iodides or bromides using palladium(II)⁴¹ or palladium(0)⁴² and copper iodide resulting in the formation of disubstituted alkynes are well-documented. Initial attempts to arylate the terminal acetylenic group of 33b with 1 equiv of iodobenzene, 0.02 equiv of (PPh₃)₂PdCl₂, and 0.005 or 0.02 equiv of CuI in Et₃N (nondeoxygenated) under N₂ at room temperature according to Hagihara's procedure,^{41a} resulted in the formation of greenish-blue reaction solutions without the expected precipitate, Et_3N ·HI. At the end of the reactions, only unreacted 33b was isolated. The arylation was further investigated with the model terminal acetylene 33a by using the same reaction conditions as above (0.02)equiv of CuI) with nondeoxygenated solvent. This time the reaction solution was brown in color, and the expected precipitate was observed. At the end of the reaction, a 92% crude yield of the desired arylated product was obtained. The presence of the greenish-blue color might indicate oxidation of the active copper(I) to the inactive copper(II) in the nondeoxygenated solvent. Without the active copper(I) catalyst, the arylation will not occur. For the model arylation reaction, on the contrary, the color of the reaction and the high yield of the desired product suggest that the oxidation of copper(I) to copper(II) did not occur. One possible explanation for the difference between the nucleoside 33b and the model compound 33a toward the arylation reaction is that the heterocyclic adenine moiety of 33b might catalyze the oxidation of copper(I) to copper(II) in the oxygen-containing solvent. When 33b was treated with 1.07 equiv of iodobenzene, 0.04 equiv of (PPh₃)₂PdCl₂, and 1.01 equiv of CuI in fully deoxygenated Et₃N at 50 °C under argon for 3 h, followed by overnight at room temperature, the disubstituted arvlalkyne nucleoside 35 was smoothly produced in 78% vield. The use of noncatalytic amounts of CuI is due to the concern of any possible interactions between the copper catalyst and the carbonyl oxygen of the N^6 -benzoyladenine moiety of 33b.43

Proceeding further along in Scheme IV, debenzoylation of 35 with methanolic ammonium hydroxide at room temperature for 18 h gave the debenzoylated nucleoside 36 in 94% yield. Desilylation of 36 with tetrabutylammonium fluoride in THF with 1% HOAc produced the hydroxy nucleoside 37 in 82-97% yield. The addition of 1% HOAc to the reaction solution serves to prevent the possible formation of ring closure products, such as 40 and 41.44 Treatment of 37 with methyltriphenoxyphosphonium iodide in CH₂Cl₂ furnished the iodo nucleoside 38 in 77-86% yield. Displacement of the iodo group of 38 proceeded smoothly with the carbanion of the

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⁽³⁹⁾ Treatment of 30a with CBr_4 (2 equiv), PPh_3 (2 equiv), and Zn dust (2 equiv) in CH_2Cl_2 according to the Corey and Fuchs procedure produced the expected dibromo olefin 32a in 95% yield. The transformation of the transformation o mation of the dibromo olefin into the corresponding terminal acetylene 33a (eq 3) using 2 equiv of n-BuLi in THF was achieved in 99% yield. Details on the conversion of 30a to 33a are found in the supplementary material.

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⁽⁴³⁾ In more recent studies, when the debenzoylated acetylenic nu-cleoside 33c was treated with 2,5-disubstituted iodobenzenes, only 0.08 equiv of CuI was needed to achieve 95–99% yields of the desired products (E. K. Yau and J. K. Coward, unpublished results).
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benzylidene-derived methyl glycinate in THF/HMPA using conditions similar to those described previously for the synthesis of 13. Acid hydrolysis of crude 39 with 80%formic acid followed by base saponification with K_2CO_3 in aqueous MeOH afforded crude 4. Purification was achieved on preparative reverse-phase (C-18) HPLC, which gave pure 4 as a white solid in 70% overall yield from 38. The structure of 4 was confirmed by spectral characterization including ¹H NMR, ¹³C NMR, FAB, and high-resolution desorption CI mass spectra.

In preliminary studies, the inhibition of COMT by 4 and 15 has been compared with the known potent product inhibitor, AdoHcy. At saturating concentrations of both substrates used, 3,4-dihydrobenzoic acid (2 mM) and AdoMet (100 μ M), 94% inhibition was observed at 90 μ M AdoHcy whereas 83 μ M of the carbon analogue, 15, produced only 23% inhibition. Similarly, at a concentration of 175 μ M of 4, only 10% inhibition of the COMT-catalyzed reaction was observed. Based on our previous experience with multisubstrate adduct inhibitors of aminopropyltransferases,⁸⁻¹¹ we expected that molecules such as 4 and 15, lacking the key recognition elements for forming the enzyme-substrate complex, would be poor inhibitors. More complex molecules such as 5 and 6, which contain these elements, should be more potent inhibitors of this important enzyme.

Conclusion

Of the two routes, A and B, which were investigated for the synthesis of 4, only route B is satisfacotry for our purposes. Route A is less efficient than route B in terms of the total steps (13 vs 10), yields, and also the formation of the overreduced olefin, most notably in the nucleoside series. The synthesis of 4 has been effected in 10 steps from the starting lactone nucleoside 18b with an overall yield as high as 18% using route B. The methods developed in this approach for the synthesis of 4 have proven to be practical and efficient for the synthesis of more highly substituted intermediates enroute to 5 and 6 (M. Burns, E. K. Yau, and J. K. Coward, unpublished results). Efforts are currently underway in our laboratory to use these new methods for the synthesis of other members of this class of compounds containing surrogates for the nucleophilic substrates of other S-adenosylmethionine dependent methyltransferases which are closely linked in cellular metabolism, e.g., phenethanolamine N-methyltransferase in catecholamine biosynthesis. Finally, the use of a linear acetylenic bond linking two substrates together as synthesized in this study, can also be applied to the possible synthesis of multisubstrate analogue inhibitors for other methyltransferases, such as bacterial protein methyltransferase or viral mRNA (guanine-7)-methyltransferase.

Experimental Section

Materials and Methods. Bis(triphenylphosphine)palladium(II) dichloride and sodium *p*-toluenesulfinate were purchased from Alfa Chemical Co. Dimethyl (phthalimidomethyl)phosphonate was purchased from Lancaster Synthesis Ltd. All reaction solvents were purified by known methods.⁴⁵ The phrase

"worked up in the usual manner" refers to washing the organic layer with water and saturated brine, drying it over MgSO₄, filtering, and evaporating to dryness. Solvent deoxygenation was accomplished by bubbling a stream of argon through the solvent during two freeze-thaw cycles using liquid nitrogen. Melting points were determined in open capillary tubes on a Thomas-Hoover Mel-Temp apparatus and are uncorrected. NMR spectra were recorded on Varian T-60, Perkin-Elmer FTR-600, Varian XL-200, and Bruker (WP-100, WP-270, WM-300, and WM-360) instruments. Proton and carbon resonances are reported as parts per million versus internal standards: tetramethylsilane for organic solvents and 3-(trimethylsilyl)-1-propanesulfonic acid, sodium salt hydrate for D₂O. Phosphorus resonances are reported relative to phosphoric acid. IR spectra were recorded on a Perkin-Elmer 298 spectrometer. UV spectra were recorded on a Perkin-Elmer 552 spectrometer. Low- and high-resolution mass spectra were recorded on a Hewlett-Packard 5987A and a VG Analytical 70-250-S instrument, respectively. Analytical thin-layer chromatography was carried out on silica gel (EM reagents, catalog no. 5775) and reverse phase (Analtech, 250 μ m). Semipreparative and preparative silica gel chromatography was performed on precoated plates (Analtech 1000, 1500, and 2000 μ m thickness), a Harrison Research Model 7924 chromatotron (Analtech 1-, 2-, and 4-mm rotor plates), flash column,⁴⁶ and filtering-column chromatography.⁴⁷ Analytical HPLC was performed on an Altex 110A pump with a Whatman Partisil PXS ODS-2 column, a Waters 6000A solvent delivery system with a Z-module C18 cartridge, and a Rainin HXP gradient system with a Dynamax C18 column. Semipreparative and preparative HPLC were done on a Rainin HXP gradient system with Dynamax C18 columns (10 mm \times 250 mm and 21.4 mm \times 250 mm). α -Methyl- γ butyrolactone (18a) was obtained from Aldrich, and the nucleoside lactone 18b was prepared as described by Lyga and Secrist.²⁹ Benzylidine-derived methyglycinate was prepared as previously described.24

(3-(Trityloxy)propanyl)triphenylphosphonium Bromide (8). A solution of 695 mg (0.45 mL, 5 mmol) of 3-hydroxypropyl bromide, 1.86 g (5.75 mmol) of trityl bromide, 30.5 mg (0.25 mmol) of DMAP, and 607 mg (0.84 mL, 6 mmol) of Et₃N in 15 mL of dry DMF was stirred at room temperature under N₂ for 1 day. This solution was poured into an ice-water bath with stirring, and the resulting mixture was extracted with CHCl₃. The CHCl₃ layer was worked up in the usual manner to give 2.23 g of a crude oily product. This crude product was purified by filtering-column chromatography on silica gel (CHCl₃; $R_f = 0.85$) to give 1.88 g (99%) of pure crystalline 3-O-tritylpropyl bromide, mp 78-80 °C. ¹H NMR (CDCl₃): δ 7.41, 7.26 (2 m, 15 H, ArH), 3.57 (t, J = 6.7Hz, 2 H, OCH₂), 3.21 (t, J = 5.8 Hz, 2 H, CH₂Br), 2.12 (m, 2 H, CH₂). Mass spectrum (rel intensity): m/e 303 (M⁺ - C₆H₅, 15).

A solution of the 3-(trityloxy)propyl bromide (6.82 g, 17.9 mmol) and 4.72 g (18 mmol) of PPh₃ in 30 mL of dry benzene was heated at reflux temperature for 10–20 h. The desired product 8 precipitated from solution, was removed from the reaction mixture by filtration, and was washed twice with dry benzene. The combined filtrate was heated at reflux for another 10–20 h. This procedure was repeated five times, and a total of six crops of the desired 8 (7.59 g, 66%) was obtained, mp 235–238 °C dec. ¹H NMR (CDCl₃): δ 7.76, 7.27 (2 m, 30 H, ArH), 3.70 (m, 2 H, CH₂P), 3.46 (t, J = 5.7 Hz, 2 H, OCH₂), 1.88 (m, 2 H, CH₂). ³¹P NMR (CDCl₃, ppm): 24.79. Anal. Calcd for C₄₀H₃₆BrOP: C, 74.65; H, 5.64; Br, 12.42. Found: C, 74.79; H, 5.85; Br, 12.20.

 N^{6} -Benzoyl-2',3'-isopropylidene-5'-(3-(trityloxy)-1propen-1-yl)-5'-deoxyadenosine (9). A suspension of 1.53 g (3.59 mmol) of hydrated 7¹⁶ in 268 mL of dry benzene was heated at reflux temperature for 60-70 min with a Dean-Stark trap. At the end of the reflux period, the reaction mixture became clear. Benzene was removed under reduced pressure, and anhydrous 7 was obtained as a white foam. ¹H NMR (DMSO-d₆) showed it was 100% free aldehyde: δ 11.27 (br s, 1 H, NH), 9.34 (s, 1 H,

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^aReagents: (a) N₂CHP(O)(OMe)₂ (34), KOBu^t, THF; (b) PhI, (PPh₃)₂PdCl₂, CuI, Et₃N; (c) NH₄OH, MeOH; (d) Bu₄N⁺F⁻, THF/HOAc (v/v, 99/1); (e) (PhO)₃P⁺CH₃I⁻, CH₂Cl₂; (f) PhCH=NCH₂CO₂Me/LDA, THF/HMPA; (g) 80% HCO₂H; (h) K₂CO₃, MeOH/H₂O.

CHO), 8.65, 8.64 (2 s, 2 H, H₈, H₂), 8.07, 7.60 (2 m, 5 H, ArH), 6.59 (s, 1 H, H₁'), 5.54 (dd, J = 1.7, 6.3 Hz, 1 H, H₃'), 5.44 (d, J= 6.3 Hz, 1 H, H_2'), 4.82 (d, J = 1.7 Hz, 1 H, H_4'), 1.55, 1.37 (2 s, 6 H, $C(CH_3)_2$). To a stirred solution of 4.42 g (6.88 mmol) of 8 in 11.5 mL of dry THF was added dropwise 4.98 mL (7.22 mmol) of 1.45 M n-BuLi in hexane at -78 °C under nitrogen. This solution was warmed to 0 °C for 10 min and then cooled back to -78 °C. To this solution was added dropwise a solution of 3.58 mmol of the freshly prepared free aldehyde 7 in 5 mL of dry THF. The reaction mixture was stirred at -78 °C for 5 h, -20 °C for 1 h, and 0 °C for 2 h. Aqueous HOAc (50%) was added to quench the reaction. The workup procedure was done in CHCl₃ in the usual manner. The crude material was purified by filtering-column chromatography on silica gel (96:4 $CH_2Cl_2/MeOH$; $R_f = 0.51$) to give 2.38 g (96%) of cis- and trans-9 as a white foam. IR (film, cm⁻¹): 3400, 3200, 3060, 2996, 2920, 1700, 1606, 1585, 1485, 1450, 1250, 1210, 1080, 910. ¹H NMR (CDCl₃): δ 9.06 (br s, 1 H, NH), 8.85, 8.83 (2 s, 2:1 ratio, 1 H, H₈), 8.09, 8.08 (2 s, 2:1 ratio, 1 H, H₂), 8.05, 7.43 (2 m, 20 H, ArH), 6.15 (overlapping two sets of d, J = 2 Hz, 1 H, H₁'), 5.92–5.50 (m, 3 H, HC=CH, H₂'), 5.14, 4.70 (two sets of dd, 2:1 ratio, 1 H, H₃'), 4.98 (dd, 1 H, H₄'), 3.13 (m, 2 H, OCH₂), 2.50, 2.31 (2 m, 2 H, CH₂), 1.65, 1.40 (2 s, 6 H, $C(CH_3)_2$). HR mass spectrum calcd for $C_{42}H_{40}N_5O_5$ (MH⁺) m/e694.3029, obsd m/e 694.3032.

2',3'-Isopropylidene-5'-(3-hydroxyprop-1-yl)-5'-deoxyadenosine (11). A solution of 350 mg (0.5 mmol) of 9, 100 mg of 5% Pd/C, and 30 mg of PtO₂ in 12 mL of absolute EtOH and two drops of glacial HOAc was shaken in a hydrogenator at 55 psi for 2 days, at which time another fresh 30 mg of PtO₂ and six drops of HOAc were added to this mixture. The hydrogenation was continued at 60 psi for 2 days. After the removal of the catalyst and solvent, a crude mixture of 10a (minor), 10b (major), and 10c (minor) was obtained. This crude mixture was dissolved in 5 mL of dry THF, and to this solution at -78 °C was added 8 mL of liquid NH₃ (freshly distilled over sodium). To this solution was added 34 mg of lithium. The reaction mixture was stirred at -78 °C for 3 h under argon and then quenched with MeOH. The mixture was worked up in CHCl₃ in the usual manner. The crude product obtained was purified by filteringcolumn chromatography on silica gel (step gradient elution with 97:3, 95:5, and finally 90:10 $CH_2Cl_2/MeOH$) to give 53 mg (30%) of the desired 11. IR (film, cm^{-1}): 3345, 3185 (very strong, OH, NH₂), 2984, 2920, 2864, 1640, 1595, 1472, 1420, 1375, 1325, 1210, 1075, 910, 865, 730. UV (MeOH): $\lambda_{max} = 258$ nm. ¹H NMR (CDCl₃): δ 8.35, 7.91 (2 s, 2 H, H₈, H₂), 6.29 (br s, 2 H, NH₂), 6.05 $(d, J = 2.3 \text{ Hz}, 1 \text{ H}, \text{H}_{1}'), 5.51 (dd, J = 2.3, 6.4 \text{ Hz}, 1 \text{ H}, \text{H}_{2}'), 4.83$ $(dd, J = 3.7, 6.4 Hz, 1 H, H_3'), 4.19 (m, 1 H, H_4'), 3.61 (t, J = 6)$ Hz, 2 H, OCH₂), 2.80 (br s, 1 H, OH), 1.75-1.39, 1.61, 1.39 (m, 2 s, 12 H, H₅', CH₂, C(CH₃)₂). ¹³C NMR (CDCl₃, ppm): 156.07 $\begin{array}{c} (C_4), \ 153.10 \ (C_2), \ 149.36 \ (C_6), \ 139.69 \ (C_8), \ 120.06 \ (C_5), \ 114.55 \\ (C(CH_3)_2), \ 90.40, \ 86.98, \ 84.29, \ 84.13 \ (C_1', \ C_2', \ C_3', \ C_4'), \ 61.92 \\ (OCH_2), \ 33.17 \ (CH_2), \ 32.36 \ (C_5'), \ 27.22, \ 25.48 \ (C(CH_3)_2), \ 22.00 \end{array}$ (CH₂). HR mass spectrum calcd for $C_{16}H_{24}N_5O_4$ (MH⁺) m/e350.1828, obsd m/e 350.1824.

2',3'-Isopropylidene-5'-(3-iodoprop-1-yl)-5'-deoxyadenosine (12). Methyltriphenoxyphosphonium iodide (427 mg, 0.95 mmol) was weighed in a glovebag under nitrogen into a two-neck round-bottom flask. To this flask at -78 °C was added dropwise a solution of 220 mg (0.63 mmol) of 11 in 12 mL of dry CH₂Cl₂ (precooled to -70 °C) under nitrogen. The reaction mixture was stirred while warming from -78 °C to room temperature over 30 min, and then stirred at room temperature for 2 h. The reaction mixture was diluted with CHCl₂ and then washed with aqueous $Na_2S_2O_3$. The organic layer was worked up in the usual manner to give 561 mg of a crude product. This crude product was immediately purified by flash column chromatography on silica gel (70:25:5 EtOAc/hexane/EtOH: $R_{t} = 0.24$) to give 218 mg (75%) of the desired 12 as a pale yellow foam. IR (film, cm^{-1}): 3320, 3168, 2982, 2940, 1640, 1595, 1472, 1420, 1370, 1325, 1292, 1210, 1155, 1080, 910, 868, 730. UV (MeOH): $\lambda_{max} = 258$ nm. ¹H NMR (CDCl₃): δ 8.37, 7.91 (2 s, 2 H, H₈, H₂), 6.33 (br s, 2 H, NH₂), 6.05 (d, J = 2.3 Hz, 1 H, H₁'), 5.52 (dd, J = 2.3, 6.4 Hz, 1 H, H₂'), 4.86 $(dd, J = 3.7, 6.4 Hz, 1 H, H_3'), 4.16 (m, J = 3.2, 3.7 Hz, 1 H, H_4'),$ $3.13 (t, J = 7 Hz, 2 H, ICH_2), 1.80-1.30, 1.62, 1.40 (m, 2 s, 12 H, 1.80-1.30)$ H_5' , CH_2 , $C(CH_3)_2$). ¹³C NMR (CDCl₃, ppm): 155.94 (C₄), 153.14 (C₂), 149.47 (C₆), 139.92 (C₈), 120.38 (C₅), 114.60 (C(CH₃)₂), 90.39, 86.77, 84.30, 84.09 (C_1' , C_2' , C_3' , C_4'), 33.05 (CH_2), 32.25 (C_5'), 27.25, 26.51 ($C(CH_3)_2$), 25.50 (CH_2), 5.97 (ICH_2). HR mass spectrum calcd for $C_{16}H_{23}IN_5O_3$ (MH⁺) m/e 460.0846, obsd m/e 460.0855.

5'-(4-Amino-4-carboxybut-1-yl)-5'-deoxyadenosine (15). To 1 equiv of LDA (generated from 1 equiv of diisopropylamine in 17.7 mL of dry THF and 1.4 mL of dry HMPA with 1.05 equiv of n-BuLi at -78 °C) was added dropwise 157 mg (0.89 mmol) of benzylidene-derived methylglycinate²⁴ at -78 °C under nitrogen. This solution was stirred at -78 °C for 30 min, at which time a solution of 388 mg (0.84 mmol) of 12 in 0.8 mL dry THF was added dropwise at the same temperature. The reaction mixture was stirred at -78 °C for 3 h, -78 °C to 0 °C over 1 h, 0 °C for 1 h, and then room temperature for 2 h. The reaction mixture was diluted with Et₂O, and the Et₂O layer was washed with ice-cold aqueous NH₄Cl. The separated organic layer was worked up in the usual manner to give 420 mg (93%) of a crude yellow foam 13. IR (film, cm⁻¹): 3212, 3180 (NH₂), 1732 (C=O). ¹H NMR (CDCl₃): δ 8.34, 7.92 (2 s, H₈, H₂), 8.26 (s, N=CH), 7.76, 7.43 (2 m, ArH), 6.39 (br s, NH₂), 6.03 (d, H₁'), 5.46 (dd, H₂'), 4.80 (dd, H₃'), 4.14 (m, H₄'), 3.96 (m, CH), 3.73 (s, OCH₃), 2.00–1.30, 1.59, 1.37 (m, 2 s, H_5' , CH_2 , $C(CH_3)_2$). Mass spectrum (rel intensity): m/e 509 (MH⁺, 3.2), 477 (M⁺ – OCH₃, 0.02), 449 (M⁺ – CO₂Me, 1.5). A solution of 370 mg (0.73 mmol) of crude 13 in 7.4 mL of 80% HCOOH was stirred at room temperature for 5 h, and the completion of the reaction was checked by TLC (reverse phase, 6:4 MeOH/H₂O). Formic acid was removed in vacuo at room temperature to give the crude methyl ester 14. The crude 14 could be purified by reverse-phase HPLC with gradient elution of 10-60% solvent A (6:4 CH_3CN/H_2O) in solvent B (0.1 M NH_4HCO_3). Pure 14 could be collected from those fractions with k' = 2.53. UV (MeOH): $\lambda_{\rm max}$ = 258 nm. ¹H NMR (CD₃OD): δ 8.22, 8.20 (2 s, 2 H, H₈, H₂), 5.94 (d, J = 4.5 Hz, 1 H, H₁'), 4.72 (overlapping two sets of dd, J = 4.5, 5 Hz, 1 H, H_2'), 4.14 (overlapping dd, J = 5 Hz, 1 H, $H_{3'}$), 3.98 (m, J = 5, 5.2 Hz, 1 H, H₄'), 3.67 (s, 3 H, OCH₃), 3.43 (t, J = 6 Hz, 1 H, CH), 1.78–1.40 (m, 8 H, CH₂). ¹³C NMR (CDCl₃, ppm): 177.21 (C=O), 157.39 (C_4) , 153.93 (C_2) , 150.70 (C_6) , 141.41 (C_8) , 120.65 (C_5) , 90.12 (C_1') , 85.64 (C4'), 75.08 (CH), 54.98, 52.41 (C2', C3'), 34.38, 35.44, 26.67, 26.37 (CH₂). FAB mass spectrum (rel intensity): m/e 381 (MH⁺, 12).

Routinely, the crude methyl ester 14 was used directly in the following base hydrolysis reaction without further purification. Crude 14 obtained as described above was dissolved in 6.7 mL of 50% aqueous MeOH. Powered K₂CO₃ (436 mg, 5 equiv) was added to the stirred solution. The reaction mixture was stirred overnight at room temperature and then neutralized with 1 N HCl. After solvent removal, the crude residue was extracted with MeOH (5×1 mL). The MeOH extract was concentrated in vacuo to give 405 mg of a crude yellow foam. Part of this foam (100 mg) was purified by preparative reverse-phase HPLC with gradient elution of 0-50% solvent A (6:4 CH₃CN/H₂O) in solvent B (0.1 M NH_4HCO_3) to give 14.7 mg of pure 15 (22%) obtained from those fractions with k' = 2.59, and 6.5 mg of the methyl ester 14 (9.5%) recovered from those fractions with k' = 6.59. For the desired 15, a colorless solid after removal of all NH₄HCO₃. UV (MeOH): $\lambda_{max} = 258 \text{ nm}$. ¹H NMR (CD₃OD): $\delta 8.22, 8.21, 8.20$ $(3 \text{ s}, 2 \text{ H}, \text{H}_8, \text{H}_2), 5.94 \text{ (d}, J = 4.8 \text{ Hz}, 1 \text{ H}, \text{H}_1'), 4.71 \text{ (overlapping })$ dd, 1 H, H₂'), 4.15 (overlapping dd, 1 H, H₃'), 4.01 (m, 1 H, H₄'), 3.52 (m, 1 H, CH), 1.81, 1.50 (2 m, 8 H, (CH₂)₄). ¹³C NMR (CD₃OD-H₂O, ppm): 175.13 (C=O), 156.93 (C₄), 153.83 (C₂),

150.20 (C₆), 141.08 (C₈), 120.11 (C₅), 89.00 (C₁'), 85.52, 85.47 (C₄'), 74.94 (CH), 74.61 (C₃'), 56.10 (C₂'), 33.89, 33.84, 31.80, 31.77, 26.01, 25.96, 25.62, 25.52 (CH₂). FAB mass spectrum (rel intensity): m/e 367 (MH⁺, 8.9). HR mass spectrum calcd for C₁₄H₂₃N₆O₃ (MH - CO₂)⁺ m/e 323.1832, obsd m/e 323.1827.

p-Methoxybenzyl Tolyl Sulfone (16). To a solution of 6.06 g (38.8 mmol) of p-methoxybenzyl chloride in 58 mL of dry DMF was added 7.28 g (40.9 mmol) of sodium p-toluenesulfinate at one portion. The reaction mixture was heated at 70 °C for 3 h, at which time DMF was removed in vacuo. The residue was dissolved in CH₃Cl and worked up in the usual manner to give 10.6 g of a white solid. This solid was recrystallized from MeOH to give 9.20 g (86%) of crystalline 16, mp 121–124 °C. ¹H NMR (CDCl₃): δ 7.54–6.77 (four sets of d, 8 H, ArH), 4.23 (s, 2 H, CH₂), 380 (s, 3 H, OCH₃), 2.42 (s, 3 H, CH₃). Anal. Calcd for C₁₅H₁₆O₃S: C, 65.19; H, 5.84. Found: C, 65.11; H, 5.78.

2-Methoxy-5-methylbenzyl Tolyl Sulfone (17). Use of the procedure described for 16 provided 5.59 g (82%) of crystalline 17, mp 128–129.5 °C. ¹H NMR (CDCl₃): δ 7.53–6.54 (m, 7 H, ArH), 4.39 (s, 2 H, CH₂), 3.33 (s, 3 H, OCH₃), 2.40, 2.27 (2 s, 6 H, CH₃). Anal. Calcd for C₁₆H₁₈O₃S: C, 66.18; H, 6.25; S, 11.04. Found: C, 66.74; H, 6.27; S, 11.34.

General Procedure for the Synthesis of 23a-d (Scheme III). Thionyl chloride (2.22 mL, 30 mmol) was added to 1.5 mmol of the carboxylic acids, $21a^{30}$ or $21b^{29}$ at 0 °C under N₂. The mixture was stirred at room temperature for 1 h. The excess of thionyl chloride was removed by a stream of nitrogen. The resulting residue was further evaporated in vacuo for 2 h to give the acid chlorides 22a or 22b, which were used directly in the following procedure. To a mechanically stirred 0.67 M solution of 16 or 17 in dry THF was added dropwise a solution of 2.09 equiv of *n*-BuLi in hexane under nitrogen from -10 to -60 °C. The suspension was stirred at -60 °C for 1 h and then cooled to -78 °C. A solution of the acid chloride, 22a (0.87 equiv) or 22b (0.5 equiv), in a minimum amount of THF was added dropwise to this orange suspension. The mixture was stirred at -78 °C for 1-3 h and -20 °C for 30 min and then guenched with aqueous HOAc. The resulting solution was adjusted to pH 7 and then partitioned between water and EtOAc. The organic layer was washed with saturated NaHCO₃ and worked up in the usual manner to give the crude products, which were purified as described below. Details on the purification and properties of 23a and 23c, both derived from 21a, are found in the supplementary material.

2',3'-Isopropylidene-5'-(1-*p*-anisoyl-1-(tolylsulfonyl)-2oxo-5-((*tert*-butyldiphenylsilyl)oxy)pent-3-yl)-5'-deoxyadenosine (23b). The crude product obtained was purified by filtering-column chromatography on silica gel (95:5 CH₂Cl₂/ MeOH; $R_f = 0.25$) to give 203 mg (76%) pure 23b as a colorless foam. IR (film, cm⁻¹): 3310, 3175 (br s, NH₂), 1714 (C==0), 1320, 1150 (SO₂). ¹H NMR (CDCl₃): δ 8.48–7.92 (m, 2 H, H₈, H₂), 7.66–6.68 (m, 18 H, ArH), 6.06 (m, 3 H, H₁', NH₂), 5.52, 5.50, 5.19 (three sets of dd, J = 2.6, 6.6 Hz, 1 H, H₂'), 5.42, 5.37, 5.33 (3 s, 1 H, R and S SO₂CH), 5.26, 4.95, 4.62 (dd, 2 m, J = 2.9, 6.6 Hz, 1 H, H₃'), 4.62, 4.32 (2 m, 1 H, H₄'), 3.73 (m, 3 H, OCH₃), 3.80–3.10 (m, 3 H, OCH₂, R and S CH), 2.38 (m, 3 H, CH₃), 2.38–1.44 (m, 4 H, H₅', CH₂), 1.66–0.88 (br m, 15 H, C(CH₃)₂, *tert*-butyl). Anal. Calcd for C₄₈H₅₅N₅O₈SSi: C, 64.77; H, 6.23; N, 7.87; S, 3.60. Found: C, 64.69; H, 6.28; N, 7.83; S, 3.55.

2',3'-Isopropylidene-5'-(1-(2-methoxy-5-methylphenyl)-1-(tolylsulfonyl)-2-oxo-5-((tert-butyldiphenylsilyl)oxy)pent-3-yl)-5'-deoxyadenosine (23d). THF and HMPA (9:1) was used as the reaction solvent. The crude product obtained was purified by filtering-column chromatography on silica gel (70:25:5 EtOAc/hexane/EtOH; $R_f = 0.23$) to give 435 mg (48%) of 23d as a foam. IR (film, cm⁻¹): 3330, 3180 (NH₂), 1716 (C=O), 1323, 1150 (SO₂). UV (MeOH): $\lambda_{max} = 258 \text{ nm}$. ¹H NMR (CDCl₃): $\delta 8.38, 8.27, 8.22, 8.01, 7.92$ (5 s, 2 H, H₈, H₂), 7.70–7.00, 6.60 (2 m, 17 H, ArH), 6.10, 5.86 (m, d, J = 2 Hz, 4 H, NH₂, CHSO₂, H₁'), $5.44, 5.37, 5.14 (dd, d, dd, J = 2, 6.7 Hz, 1 H, H_2'), 5.22, 4.90, 4.54$ $(dd, 2 m, J = 2.5, 6.7 Hz, 1 H, H_3'), 4.54, 4.18 (2 m, 1 H, H_4'), 3.64$ (m, 2 H, OCH₂), 3.34, 3.30, 3.26 (3 s, 3 H, OCH₃), 3.07 (m, 1 H, CH), 2.35, 2.26, 2.14 (3 s, 3 H, CH₃), 2.20–1.25 (m, 10 H, H₅', CH₂, C(CH₃)₂), 0.93, 0.89, 0.87 (3 s, 9 H, tert-butyl). ¹³C NMR (CDCl₃, ppm): 203.08, 202.85, 202.70 (C=O), 155.68, 155.53, 149.29, 144.36, 144.29, 135.45, 135.38, 135.31, 133.60, 133.48, 131.27, 129.77, 129.67, 129.58, 129.51, 128.80, 128.68, 128.63, 127.72, 127.64, 127.59, 127.55,

110.63 (Ar), 155.19, 155.13 (C₄), 153.14, 153.02 (C₂), 140.07, 139.47 (C₈), 135.15, 135.13 (C₆), 120.17, 120.15 (C₅), 115.81, 115.06, 114.82 (C(CH₃)₂), 90.08, 90.02, 89.43 (C₁'), 84.28, 84.25, 84.07, 83.84, 83.68, 83.09 (C₂', C₃', C₄'), 70.05, 69.93 (CHSO₂), 60.72 (OCH₂), 55.52, 55.39 (OCH₃), 46.38, 46.06, 45.63 (CH), 33.08, 33.01, 32.33, 31.94, 31.34 (C₅', CH₂), 25.44, 25.38 (*tert*-butyl, C(CH₃)₂), 21.54, 20.41 (ArCH₃), 19.07, 18.98 (*tert*-butyl). FAB mass spectrum (rel intensity): m/e 904 (M⁺, 33), 614 (M⁺-289, 6).

2',3'-Isopropylidene-5'-(1-p-anisoyl-1-(tolylsulfonyl)-2-((diphenoxyphosphinyl)oxy)-5-((tert-butyldiphenylsilyl)oxy)-2-penten-3-yl)-5'-deoxyadenosine (24b). A solution of 589 mg (0.66 mmol) of 23b and 237 mg (0.26 mL; 1.99 mmol) of N,N-dimethylformamide dimethyl acetal in 1.33 mL of dry DMF was stirred at room temperature under nitrogen for 10 h. DMF was removed in vacuo, and the resulting residue was coevaporated twice with toluene to give 588 mg of crude 23e. This crude 23e, 235 mg (0.18 mL, 0.88 mmol) of diphenyl phosphochloridate, and 18.2 mg (0.14 mmol) of DMAP were dissolved in 1.1 mL of dry CH₃CN. To this solution was added 0.144 mL (1.03 mmol) of $\ensuremath{\text{Et}}_3N$ at room temperature under nitrogen. The reaction mixture was heated at 64 °C for 20 min. The cooled solution was diluted with CHCl₃ and worked up in the usual manner to give 627 mg of crude 24e. This crude product was dissolved in a solution of 23.6 mL of $HOAc/EtOH/H_2O$ (2:3:1, v:v:v). This mixture was stirred at room temperature for 5-6 h. After the removal of solvent in vacuo, the resulting residue was purified by filtering-column chromatography on silica gel (65:30:5 EtOAc/hexane/EtOH; R_f = 0.4) to give 286 mg (38%) of the desired 24b and 83 mg (11%) recovery) of the N⁶-protected 24e. For 24b. IR (film, cm⁻¹): 3320, 3175 (br s, NH₂), 1320, 1150 (SO₂). ¹H NMR (CDCl₃): δ 8.34, 8.32, 8.23, 8.21, 7.99, 7.79 (ms, 2 H, H₈, H₂), 7.70-6.36 (m, 28 H, ArH), 6.11-5.84 (m, 3 H, H₁', NH₂), 5.48-5.29 (three sets of dd, 1 H, $H_{2'}$), 5.50, 4.84, 4.55 (2 m, 1 H, $H_{3'}$), 4.55, 4.44, 4.16 (m, 1 H, H₄'), 4.04, 2.70 (2 m, 1 H, R and S CH), 3.82-3.36 (m, 5 H, OCH₃, OCH₂), 2.38, 2.34, 2.32, 2.29 (4 s, 3 H, ArCH₃), 2.20-1.36 (m, 10 H, $H_{5'}$, CH_{2} , $C(CH_{3})_{2}$), 1.06, 1.04, 0.89, 0.83 (4 s, 9 H, tert-butyl). ³¹P NMR (CDCl₃, ppm): -19.65, -19.94, -20.15, -20.37. FAB mass spectrum (rel intensity): m/e 1122 (MH⁺, 71). HR mass spectrum calcd for $C_{60}H_{65}N_{50}O_{11}PSSi$ (MH⁺) m/e1122.3908, obsd m/e 1122.3943.

N⁶-Benzoyl-2',3'-isopropylidene-5'-(2-hydroxytetrahydrofuran-3-yl)-5'-deoxyadenosine (29b). To a solution of 1.42 g (3 mmol) of 18b²⁹ in 40 mL of dry toluene and 30 mL of dry THF was added dropwise 10.1 mL of 1 M DIBAL in toluene at -78 °C under nitrogen. The reaction mixture was stirred at -78 °C for 3 h. A solution of 3 mL of MeOH and 2 mL of aqueous NH_4Cl was added to quench the reaction. The mixture was extracted with EtOAc, and the EtOAc extract was worked up in the usual manner to give 1.35 g of a crude foam. This crude foam was purified by filtering-column chromatography on silica gel (98:2 $CH_2Cl_2/MeOH$) to give 1.06 g (74%) of foamy 29b (100% lactol form). IR (film, cm⁻¹): 3265 (OH), 2980, 2930, 1700, 1610, 1570, 1510, 1453, 1250, 1210, 1080. ¹H NMR (CDCl₃): δ 9.04 (br s, 1 H, NH), 8.83, 8.11 (2 s, 2 H, H₈, H₂), 8.04, 7.57 (2 m, 5 H, ArH), 6.11 (m, 1 H, H_1'), 5.52 (m, 1 H, H_2'), 5.37, 5.21, 5.07 (3 s, 1 H, OCH), 4.92 (m, 1 H, H₃'), 4.30 (m, 1 H, H₄'), 4.08-3.66 (m, 2 H, OCH₂), 3.09, 2.54 (2 m, 1 H, R and S CH), 2.18-1.92 (m, 4 H, H₅', CH₂), 1.62, 1.40 (2 s, 6 H, C(CH₃)₂). ¹³C NMR (CDCl₃, ppm): 164.78 (N-C=O), 152.66 (C₂), 151.40 (C₄), 149.96 (C₆), 142.28 (C₈), 133.86, 132.63, 128.75, 127.97 (Ar), 123.83 (C₅), 114.97 (C- $(CH_3)_2$, 102.87, 97.81 (OCH), 90.59 (C_1') , 85.80 (C_4') , 84.49 (C_2') , 84.17 (C₃'), 66.87, 66.61 (OCH₂), 43.46, 41.14 (CH), 35.65, 32.58 (CH₂), 30.89, 29.19 (C₅'), 27.22, 25.44 (C(CH₃)₂). FAB mass spectrum (rel intensity): m/e 482 (MH⁺ 2.53), 464 (MH⁺ - H₂O, 3.21). HR mass spectrum calcd for $C_{24}H_{28}N_5O_6$ (MH⁺) m/e482.2040, obsd m/e 482.2023.

 N^6 -Benzoyl-2',3'-isopropylidene-5'-(1-formyl-3-((*tert*-butyldiphenylsilyl)oxy)-1-propyl)-5'-deoxyadenosine (30b). To a solution of 481 mg (1 mmol) of **29b** and 68.1 mg (1.01 mmol) of imidazole in 2.81 mL of dry DMF was added dropwise 0.26 mL (d = 1.06, 1 mmol) of *tert*-butyldiphenylchlorosilane at room temperature under nitrogen. The solution was stirred at 50 °C for 40 min, at which time 34 mg of imidazole (0.5 mmol) was added in one portion and then immediately followed by dropwise addition of 0.13 mL of *tert*-butyldiphenylchlorosilane (0.5 mmol). The resulting mixture was allowed to continue to stir at 50 °C for 40 min. The above additions and stirring procedure were repeated again. At the end of the reaction, DMF was removed in vacuo, and the resulting residue was worked up in CHCl₃ in the usual manner to give 751 mg of crude syrup. This syrup was purified by filtering-column chromatography on silica gel (8:1:1 CHCl₃/acetone/EtOAc; $R_f = 0.29$) to give 636 mg (88%) of a ca. 3:1 mixture of the desired **30b** and the O-silylated lactol **31b**, isolated as a foam. This material is suitable for use in further transformations.

Pure 30b (free of 31b) could be obtained in 32-39% yield when DMF/HMPA (1:2 or 1:1; v:v) was used as the reaction solvent in the procedure described above. IR (film, cm⁻¹): 3240 (NH), 3060, 2923, 2844, 1700 (C=O), 1605, 1580, 1505, 1450, 1296, 1210, 1090, 700. ¹H NMR (CDCl₃): δ 9.57, 9.53 (two sets of d, J = 1.2Hz, 1 H, R and S CHO), 9.00 (br s, 1 H, NH), 8.89, 8.06 (2 s, 2 H, H₈, H₂), 8.04, 7.64, 7.44 (2 m, 15 H, ArH), 6.07 (d, J = 2 Hz, $1 \text{ H}, \text{H}_{1}$), 5.46 (dd, $J = 2, 6 \text{ Hz}, 1 \text{ H}, \text{H}_{2}$), 4.90 (dd, $1 \text{ H}, \text{H}_{3}$), 4.30 (m, 1 H, H₄'), 3.65 (m, 2 H, OCH₂), 2.67 (m, 1 H, CH), 2.20, 1.84 (2 m, 4 H, H_5' , CH₂), 1.62, 1.39 (2 s, 6 H, C(CH₃)₂), 1.02, 0.99 (2 s, 9 H, *tert*-butyl). ¹³C NMR (CDCl₃, ppm): 203.38, 203.30 (*R* and S CHO), 164.65 (C=O), 153.77 (C2), 151.17 (C4), 149.80 (C6), 142.34 (C₈), 135.52, 133.31, 133.26, 132.78, 129.75, 129.64, 129.03 (Ar), 123.70 (C₅), 115.01 ($C(CH_3)_2$), 90.34, 90.25 (C₁'), 85.70–83.90 (ms, C_2' , C_3' , C_4'), 61.15 (OCH₂), 45.75 (CH), 32.00 (C_5'), 27.02, 25.43 ($C(CH_3)_2$), 26.74, 19.07 (tert-butyl). Anal. Calcd for C40H45N5O6Si: C, 66.74; H, 6.30; N, 9.73. Found: C, 66.70; H, 6.41; N, 9.65.

N⁶-Benzoyl-2',3'-isopropylidene-5'-(1,1-dibromo-5-((tertbutyldiphenylsilyl)oxy)-1-penten-3-yl)-5'-deoxyadenosine (32b). A suspension of 635 mg (2.42 mmol) of PPh₃, 803 mg (2.42 mmol) of CBr₄, and 158 mg (2.42 mmol) of Zn dust in 19.5 mL of dry CH₂Cl₂ was stirred at room temperature under nitrogen for 1 day, at which time a solution of 290 mg (0.40 mmol) of 30b and 31b (3:1) in 1 mL of CH_2Cl_2 was added. The reaction mixture was stirred at room temperature for 7 h. Solvent was removed under reduced pressure, and the resulting residue was purified by filtering-column chromatography on silica gel (65:30:5 Et-OAc/hexane/EtOH) to give 86 mg of pure 32b ($R_f = 0.6$), in addition to a mixture of 128 mg of 32b and silvlated lactol (31b) from the starting material ($R_f = 0.52$). This mixture was further purified by preparative TLC (eluted three times with 97:3 $CHCl_3/MeOH$) to give 65 mg of pure 32b and 48 mg of a 1:1 mixture of 32b and of 31b. The total yield of the desired 32b was 175 mg (50%). The yield of 32b, corrected for the amount of unreactive 31b in the starting material, is 66% (49.6 ÷ 0.75). IR (film, cm⁻¹): 3240, 3065, 2938, 2858, 1700, 1605, 1580, 1450, 1250, 1210, 1155, 1100, 867, 755, 700. ¹H NMR (CDCl₃): δ 9.24 (br s, 1 H, NH), 8.82, 8.79 (2 s, 1 H, H₈), 8.09 (s, 1 H, H₂), 8.02, 7.40 (2 m, 15 H, ArH), 6.13 (two sets of d, J = 10 Hz, 1 H, HC==), 6.10 (d, J = 2 Hz, 1 H, H₁'), 5.54 (m, 1 H, H₂'), 4.87 (m, 1 H, H₃'), 4.26 (m, 1 H, H₄'), 3.59 (m, 2 H, OCH₂), 2.80 (m, 1 H, CH), 1.96-1.44 (m, 4 H, H₅', CH₂), 1.63, 1.39 (2 s, 6 H, C(CH₃)₂), 1.04, 1.00 (2 s, 9 H, tert-butyl). Anal. Calcd for $C_{41}H_{45}Br_2N_5O_5Si$: C, 56.23; H, 5.18; Br, 18.25; N, 8.00. Found: C, 56.16; H, 5.22; Br, 18.22; N, 7.94.

Dimethyl (Diazomethyl)phosphonate (34).40 A solution of 13.2 g (49.1 mmol) of dimethyl (phthalimidomethyl)phosphonate and 1.66 g (1.64 mL; 51.8 mmol) of anhydrous hydrazine in 84.7 mL of dry MeOH was stirred at room temperature under nitrogen for 3 days. A white precipitate was removed by filtration and washed twice with dry MeOH. The combined filtrate was evaporated at 0 °C in vacuo, and the resulting residue was immediately dissolved in 13.6 mL of cold glacial HOAc. While the mixture was mechanically stirred at -17 °C in a round-bottom flask equipped with an inner thermometer, a solution of 4.96 g (71.2 mmol) of NaNO₂ in 11.3 mL of ice-cold H₂O was added dropwise. The reaction temperature was kept below -10 °C during the addition of NaNO₂. The reaction mixture was stirred at -10°C for 30 min, 0 °C for 15 min, and then cooled back to -10 °C. To this cold mixture was added slowly, in small portions, 19.6 g of NaHCO₃. The mixture was warmed to room temperature, and another 9.82 g of NaHCO₃ was added in small portions with vigorous stirring. After the resulting foaming subsided, the mixture was filtered. The filter was washed twice with CH₂Cl₂, and the combined filtrate was diluted with 200 mL of CH₂Cl₂. The CH₂Cl₂ layer was washed with saturated NaHCO₃ and NaCl, dried over with MgSO₄, filtered, and concentrated in vacuo. A crude yellow liquid (5.48 g) was obtained. This liquid was subjected to vacuum distillation (short-path) and 3.2 mL (d = 1.27; 4.06 g; 55%) or pure 34 was collected at 45–56 °C (0.15–0.3 Torr) (lit.⁴⁰ bp 59 °C (0.42 Torr)). IR (film, cm⁻¹): 2110 (C=N=N), 1250 (P=O), 1030 (P-O-C). ¹H NMR (CDCl₃): δ 3.81 (d, J = 0.6 Hz, 1 H, CH), 3.79 (d, J = 11.8 Hz, 3 H, OCH₃). ¹H NMR (neat): δ 4.42 (d, J = 10.5 Hz, 1 H, CH), 3.68 (d, J = 11.8 Hz, 3 H, OCH₃). ¹³C NMR (CDCl₃, ppm, P-C coupled): 54.07 (d, J = 5.4 Hz, OCH₃), 29.64 (d, J = 230.9 Hz, CH). ³¹P NMR (CDCl₃, ppm): 22.52. Mass spectrum (rel intensity): m/e 150 (M⁺, 40).

N⁶-Benzoyl-2',3'-isopropylidene-5'-(5-((*tert*-butyldiphenylsilyl)oxy)-1-pentyn-3-yl)-5'-deoxyadenosine (33b). To a magnetically stirred solution of 64.1 mg (0.57 mmol) of potassium tert-butoxide in 1.07 mL of dry THF was added dropwise 85.7 mg (0.57 mmol) of 34 in 1.42 mL of dry THF at -78 °C under nitrogen. This solution was stirred at the same temperature for 15 min, and to this solution was added dropwise 360 mg (0.5 mmol) of a ca. 1:1 mixture of 30b and 31b in 1.42 mL of THF. The reaction mixture was stirred at -78 °C for 5 h and then guenched with aqueous HOAc. The mixture was worked up in CH_2Cl_2 in the usual manner to give 381 mg of a crude foam. This foam was purified by filtering-column chromatography on silica gel (82:10:8 $Et_2O/CHCl_3/EtOAc; R_f = 0.4$) to give 111 mg (31%) of pure 33b. The corrected yield of 33b from 30b is 56.4% ($31 \div 0.55$). IR (film, cm⁻¹): 3308 (HC=), 2119 (C=C). ¹H NMR (CDCl₃): δ 9.04 (br s, 1 H, NH), 8.84, 8.83 (2 s, 1 H, H_8), 8.11, 8.08 (2 s, 1 H, H_2), 8.04, 7.64, 7.38 (2 m, 15 H, ArH), 6.12 (d, J = 2.5 Hz, 1 H, H₁'), 5.54 (overlapping two sets of dd, J = 2.5, 6.8 Hz, 1 H, H₂'), 4.94 (m, 1 H, H₃'), 4.56 (m, 1 H, H₄'), 3.76 (m, 2 H, OCH₂), 2.80 (m, 1 H, CH), 2.07, 2.05 (2 d, J = 2.4 Hz, 1 H, R and S HC=), 1.90 (m, 2 H, H₅'), 1.64 (m, 2 H, CH₂), 1.64, 1.40 (2 s, 6 H, C(CH₃)₂), 1.03, 0.95 (2 s, 9 H, tert-butyl). ¹³C NMR (CDCl₃, ppm): 165.30, 165.24 (C==O), 153.37 (C₂), 151.86, 151.74 (C₄), 150.37 (C₆), 143.07, 142.89 (C8), 136.19, 134.25, 133.42, 130.23, 130.17, 129.46, 128.53, 128.28, 128.22, 128.19 (Ar), 124.24 (C₅), 115.42, 115.30 (C(CH₃)₂), 104.77 $(\text{HC} \equiv)$, 91.48, 91.32 (C_1') , 86.56, 86.17 (*R* and *S* \equiv C), 85.73, 85.60, 84.99, 84.94, 84.68, 84.50 (C₂', C₃', C₄'), 61.82, 61.75 (OCH₂), 39.35, 38.71, 38.46, 37.95 (C₅', CH₂), 27.84, 27.82, 27.45, 27.36, 26.06, 25.83, 25.17, 19.83, 19.73 (C(CH₃)₂, CH, tert-butyl). Mass spectrum (rel intensity): m/e 715 (M⁺, 0.12), 658 (M⁺ - C₄H₉, 10.0). Anal. Calcd for C41H45N5O5Si: C, 68.79; H, 6.34; N, 9.78. Found: C, 68.73; H, 6.44; N, 9.37.

N⁶-Benzoyl-2',3'-isopropylidene-5'-(5-((*tert*-butyldiphenylsilyl)oxy)-1-phenyl-1-pentyn-3-yl)-5'-deoxyadenosine (35). To a 15-mL two-neck round-bottom flask containing 8.11 mg (0.04 equiv) of PdCl₂(PPh₃)₂ and 55.9 mg (1.01 equiv) of CuI was added at room temperature under argon a solution of 208 mg (0.29 mmol) of 33b in 3.5 mL of fully deoxygenated Et_3N , followed by 35 μ L (d = 1.82; 0.31 mmol) of iodobenzene. The reaction mixture was heated at 50 °C with stirring for 3 h (white precipitate was formed) and then at room temperature overnight. Solvent was removed in vacuo, and the resulting residue was dissolved in CHCl₃ and washed twice with a 2.5% EGTA suspension. The CHCl₃ layer was then worked up in the usual manner to give 235 mg of a crude foam. This foam was purified by filtering-column chromatography on silica gel (9:1 Et₂O/CHCl₃; $R_f = 0.23$) to give 178 mg (78%) of a pale yellow foam 35. IR (film, cm⁻¹): 2214 (very weak, C=C). ¹H NMR (CDCl₃): δ 9.27 (br s, 1 H, NH), 8.83 (br, s, 1 H, H₈), 8.11, 8.09 (2 s, 1 H, H₂), 8.04-7.26 $(2 \text{ m}, 20 \text{ H}, \text{ArH}), 6.13 \text{ (d}, J = 2.4 \text{ Hz}, 1 \text{ H}, \text{H}_1'), 5.55 \text{ (overlapping)}$ two sets of dd, J = 2.4, 6.7 Hz, 1 H, H_2'), 5.01 (overlapping two sets of dd, J = 3.5, 6.7 Hz, 1 H, $H_{3'}$), 4.63 (m, 1 H, $H_{4'}$), 3.85 (m, 2 H, OCH₂), 2.99 (m, 1 H, CH), 1.96 (m, 2 H, H₅'), 1.80 (m, 2 H, CH₂), 1.64, 1.40 (2 s, 6 H, C(CH₃)₂), 1.05, 0.98 (2 s, 9 H, tert-butyl). ¹³C NMR (CDCl₃, ppm): 164.51 (C=O), 152.79 (C₂), 151.50 (C₄), 149.71 (C₆), 142.40, 142.19 (C₈), 135.54-127.61 (Ar), 123.66, 123.56, 123.53 (C₅, Ar-C=), 114.78, 114.61 ($C(CH_3)_2$), 91.41, 91.04 (R and $S \text{ Ar-}C \equiv$), 90.92, 90.65 (C₁'), 85.47, 85.04, 84.48, 84.33, 84.07, 83.89 (C_2', C_3', C_4') , 83.14, 82.71 (R and S = C), 61.42 (OCH₂), 39.11, 38.14 (C₅', CH₂), 27.26, 26.77, 26.13, 25.46, 19.23, 19.14 (C(CH₃)₂, CH, tert-butyl). Mass spectrum (rel intensity): m/e 792 (M⁺, 1.71), 735 ($M^+ - C_4 H_9$, 2.0).

2',3'-Isopropylidene-5'-(5-((*tert*-butyldiphenylsilyl)oxy)-1-phenyl-1-pentyn-3-yl)-5'-deoxyadenosine (36). A solution of 170 mg (0.22 mmol) of 35 in 8.7 mL of concentrated

NH4OH and 15 mL of MeOH was stirred at room temperature for 18 h. Solvent was removed in vacuo, and the resulting residue was purified by filtering-column chromatography on silica gel (initial elution with 35:65 EtOAc/CHCl₃, and then 35:63:2 Et-OAc/CHCl₃/MeOH) to give 138 mg (94%) of 36 as a colorless foam. IR (film, cm⁻¹): 3338, 3189 (NH₂), 2222 (weak, C=C). ¹H NMR (CDCl₃): δ 8.37 (s, 1 H, H₈), 7.92, 7.89 (2 s, 1 H, H₂), 7.66, 7.29 (2 m, 15 H, ArH), 6.07 (d, J = 2.4 Hz, 1 H, H_1'), 6.04 (br s, 2 H, NH₂), 5.54 (overlapping two sets of dd, J = 2.4, 6.3 Hz, 1 H, H_{2}'), 4.98 (m, 1 H, H_{3}'), 4.58 (m, 1 H, H_{4}'), 3.83 (m, 2 H, OCH₂), 2.97 (m, 1 H, CH), 2.00, 1.80 (2 m, 4 H, H₅', CH₂), 1.63, 1.39 (2 s, 6 H, C(CH₂)₂), 1.04, 0.97 (2 s, 9 H, tert-butyl). ¹³Č NMR (CDCl₃, ppm): 155.52 (C₄), 153.17, 153.08 (C₂), 149.49, 149.40 (C₆), 140.09, 139.82 (C₈), 135.54, 133.85, 133.74, 131.67, 129.53, 128.10, 127.60 (Ar), 123.66 (C₅), 120.43 (Ar-C=), 114.64, 114.43 (C(CH₃)₂), 91.58, 91.23 (Ar-C≡), 90.75, 90.39 (C₁'), 85.29, 84.88, 84.58, 84.38, 84.09, 83.89 (C_2' , C_3' , C_4'), 83.03, 82.62 (*R* and *S* ==C), 61.47 (OCH₂), 39.13, 38.19 (C_5' , CH₂), 27.28, 26.86, 26.77, 26.11, 25.48, 19.14 (*tert*-butyl, 27.28), 26.28, 26.27, 26.11, 25.48, 19.14 (*tert*-butyl, 27.28), 26.28, 26.28, 26.27, 26.21, 25.28, 26.28, 26.27, 26.21, 25.28, 26.28, 26.27, 26.21, 25.28, 26.28, 2 CH, C(CH₃)₂). Mass spectrum (rel intensity): m/e 688 (M⁺, 0.5), 673 $(M^+ - CH_3, 0.7)$, 630.5 $(M^+ - C_4H_9, 18.6)$. Anal. Calcd for C₄₀H₄₅N₅O₄Si: C, 69.84; H, 6.59; N, 10.18. Found: C, 69.93; H, 6.62; N, 10.14.

2',3'-Isopropylidene-5'-(5-hydroxy-1-phenyl-1-pentyn-3yl)-5'-deoxyadenosine (37). To a solution of 191 mg (0.28 mmol) of 36 in 3.4 mL of dry THF was added 34 µL of glacial HOAc and 0.6 mL of 1 M tetrabutylammonium fluoride in THF at room temperature under nitrogen. The progress of the reaction was monitored by TLC (95:5 CHCl₃/MeOH) and additional portions $(2 \times 0.1, 2 \times 0.06, 1 \times 0.03 \text{ mL})$ of 1 M tetrabutylammonium fluoride in THF were added at reaction intervals of 180, 270, 330, 390, and 450 min, respectively. The reaction was completed after 520 min at room temperature and the mixture was diluted with CH_2Cl_2 and water. The CH_2Cl_2 layer was worked up in the usual manner to give a crude product, which was then purified by filtering-column chromatography on silica gel (95:5 CHCl₃/MeOH) to give 121 mg (0.27 mmol; 97%) of pure 37. IR (film, cm^{-1}): 3325, 3180 (very strong br s, NH₂, OH), 2218 (weak, C=C). ¹H NMR (CDCl₃): δ 8.36, 8.35 (2 s, 1 H, H₈), 7.90, 7.88 (2 s, 1 H, H₂), 7.30 (m, 5 H, ArH), 6.06, 6.05 (d, J = 2.3 Hz, 1 H, H_1'), 5.92, 5.85 (2 br s, 2 H, NH₂), 5.61, 5.51 (two sets of dd, J = 2.3, 6.2 Hz, 1 H, $H_{2'}$), 4.98 (m, 1 H, $H_{3'}$), 4.63, 4.55 (2 m, 1 H, $H_{4'}$), 3.80 (m, 2 H, OCH₂), 2.87 (m, 1 H, CH), 2.05–1.64 (m, 4 H, H₅', CH₂), 1.63, 1.39 (2 s, 6 H, C(CH₃)₂). ¹³C NMR (CDCl₃, ppm): 155.86 (C₄), 153.13, 153.05 (C₂), 149.20, 149.05 (C₆), 140.00, 139.74 (C₈), 131.61, 131.52, 128.18, 127.80, 120.03 (Ar), 123.43, 123.37 (C₅), 114.55, 114.20 $(C(CH_3)_2)$, 91.35, 91.16 (Ar- $C \equiv$), 91.09, 90.28 (C_1') , 85.29, 85.09, 84.67, 84.19, 84.03, 83.69, 83.14, 82.93 (C_2 ', C_3 ', C_4 ', =C), 60.10, 59.94 (OCH₂), 38.98, 38.31, 37.58 (C₅', CH₂), 27.20, 26.14, 25.57, 25.40 ($C(CH_3)_2$, CH). HR mass spectrum (desorption CI) calcd for C₂₄H₂₈N₅O₄ (MH⁺) 450.2141, found 450.2126.

2',3'-Isopropylidene-5'-(5-iodo-1-phenyl-1-pentyn-3-yl)-5'deoxyadenosine (38). Crude 38 (249 mg) was obtained from 0.23 mmol of 37 by using a similar method to that described for the preparation of 12. This crude product was purified by filtering-column chromatography on silica gel (95:5 CHCl₃/2-propanol; $R_f = 0.19$) to give 111 mg (86%) of the desired 38. IR (film, cm⁻¹): 3320, 3165, 2985, 2936, 2224 (weak, C=C), 1700, 1600, 1590, 1580, 1425, 1330, 1210, 1085. ¹H NMR (CDCl₃): δ 8.38, 8.36 (2 s, 1 H, H_8), 7.92, 7.91 (2 s, 1 H, H_2), 7.30 (m, 5 H, ArH), 6.06 (d, J = 2.3Hz, 1 H, H_1'), 6.02 (br s, 2 H, NH₂), 5.57 (two sets of dd, J = 2.3, 6.3 Hz, 1 H, H₂'), 5.01 (m, 1 H, H₃'), 4.57 (m, 1 H, H₄'), 3.26 (m, 2 H, ICH₂), 2.91, 2.80 (2 m, 1 H, R and S CH), 2.00 (m, 4 H, H₅', CH₂), 1.62, 1.39 (2 s, 6 H, C(CH₃)₂). ¹³C NMR (CDCl₃, ppm): 155.91 (C₄), 153.10, 152.99 (C₂), 149.21, 149.14 (C₆), 140.05, 139.75 (C₈), 131.60, 131.53, 128.15, 127.91, 120.28, 120.17 (Ar), 123.16 (C₅), 114.53, 114.28 ($C(CH_3)_2$), 90.81, 90.31 (Ar- $C \equiv$), 89.78, 89.54 (C_1), 85.09, 84.76, 84.53, 84.22, 83.95, 83.68, 83.48 (C_2' , C_3' , C_4' , $\equiv C$), 39.01, 38.27, 37.58 (C_5' , CH_2), 27.14, 25.37, 25.31 ($C(CH_3)_2$, CH). HR mass spectrum (desorption CI) calcd for C₂₄H₂₇N₅O₃I (MH⁺) m/e 560.1159, obsd m/e 560.1180.

2',3'-Isopropylidene-5'-(6-(methoxycarbonyl)-6-(benzylideneamino)-1-phenyl-1-hexyn-3-yl)-5'-deoxyadenosine (39). A crude foam of 39 (140 mg, >100%) was obtained from 0.22 mmol of 38 by using a similar method to that described for the preparation of 13. ¹H NMR (CDCl₃): δ 8.34, 8.33 (2 s, H₈), 8.30–8.25 (5 s, N=CH), 7.88, 7.86 (2 s, H₂), 7.76, 7.42 (2 m, ArH), 6.03, 6.00 (two sets of d, J = 2.3 Hz, H_1'), 5.86 (br s, NH₂), 5.54, 5.47 (two sets of dd, J = 2.3, 6.3 Hz, H_2'), 4.95 (dd, H_3'), 4.56 (m, H_4'), 3.78 (m, CH), 2.70 (m, CH), 2.20-1.37 (m, H₅', (CH₂)₂), 1.60, 1.37 (2 s, C(CH₃)₂). Mass spectrum (desorption CI, rel intensity): m/e609 (MH⁺, 8.4).

5'-(6-Amino-6-carboxy-1-phenyl-1-hexyn-3-yl)-5'-deoxyadenosine (4). A crude sample of 4 (41.6 mg) was obtained from 140 mg of 39 by using a similar method to that described for the preparation of 15. This crude product was purified by preparative reverse-phase HPLC with gradient elution of 0-100% solvent A (6:4 CH₃CN/H₂O) in solvent B (H₂O) to give 20.6 mg (70% yield from 38) of the desired 4 as a foam. UV (MeOH): $\lambda_{max} = 251$ nm, $\lambda_{min} = 240$ nm. ¹H NMR (CD₃OD, δ): 8.25, 8.24, 8.21, 8.20 (4 s, 2 H, H₈, H₂), 7.30 (m, 5 H, ArH), 5.98 (two sets of d, 1 H, H_{1}'), 4.85 (overlapping with HOD peak, H_{2}'), 4.40 (m, 1 H, H_{4}'), 4.25 (m, 1 H, H₃'), 3.59 (m, 1 H, CH), 2.86 (m, 1 H, CH), 2.28-1.60 (m, 6 H, H₅', (CH₂)₂). ¹³C NMR (CD₃OD, ppm): 174.38 (C=O), 157.37 (C₄), 153.94 (C₂), 150.62 (C₆), 141.78, 141.49 (C₈), 132.65, 132.58, 132.65, 128.94, 120.77 (Ar), 124.95 (C₅), 92.77, 92.16, 92.09 $(Ar-C \equiv), 90.61, 90.55, 90.20 (C_1'), 84.44, 84.40 (C_4'), 83.86, 83.74,$

83.70 (=C), 75.42, 75.23 (R and S CH), 74.94, 74.90 (C₃'), 56.17, 56.13 (C₂'), 40.43, 40.11, 39.96 (C₅', CH₂), 32.36, 31.39, 31.20, 30.58, 30.47, 30.07 (CH₂, R and S CH). FAB mass spectrum (rel intensity): m/e 467 (MH⁺, 16.8). HR mass spectrum (desorption CI) calcd for $(MH - CO_2)^+ m/e C_{22}H_{27}N_6O_3 423.2145$, obsd m/e423.2139.

Acknowledgment. The research was supported by a grant from the National Cancer Institute, CA28097. We are grateful to Jane LeBlanc for her assistance in the preparation of this manuscript. We thank Mark Burns for carrying out the enzyme inhibition studies and for his critical reading of several drafts of this paper. We also thank Jim Windak for his extraordinary efforts to obtain the mass spectral data.

Supplementary Material Available: Procedures for the synthesis of 23a, 23c, 24a, 25a, 30a, 32a, and 33a with complete spectral data (3 pages). Ordering information is given on any current masthead page.

Total Synthesis and Absolute Stereochemistry of (+)-Xestoquinone and Xestoquinol

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Received November 15, 1989

The first total synthesis of (+)-xestoquinone 1, a biologically active marine natural product isolated from tropical marine sponges, was achieved. Optically pure hydroxymethyl ketone derivative (4aR, 5S, 8aR)-(+)-8 was converted to enone (+)-15 via the reactions of eight steps. The Diels-Alder reaction of (+)-15 with 3,6-dimethoxybenzocyclobutene (16) afforded an adduct (+)-17 of a tetracyclic system, which was converted to (12bS)-xestoquinone 1 via a series of five-step reactions. The CD spectrum of the synthetic sample was identical with that of the natural xestoquinone (+)-1. Therefore, the absolute stereochemistry of (+)-xestoquinone 1 was determined to be 12bS. Finally, xestoquinone (12bS)-(+)-1 was converted to xestoquinol (12bS)-2, although xestoquinol 2 itself has not been isolated yet as a natural product. The absolute configuration of xestoquinone 1 was also established by the comparison of the CD spectrum of naphthalene-diene derivative 22 with the theoretically calculated CD curve of a model compound (12bS)-23.

A variety of novel quinone and hydroquinone compounds with biological activites have been isolated from tropical marine sponges (Chart I). Xestoquinone (+)-1 was isolated from the Okinawan sponge Xestospongia sapra as a cardiotonic constituent by Nakamura.² From the same Okinawan sponge, Kitagawa³ isolated halenaquinol (+)-4 together with its sulfate ester, and hydroquinone (+)-4 was easily oxidized to give halenaquinone (+)-3, which had been originally isolated from the sponge Xestospongia exigua in Western Caroline Islands by Scheuer.⁴ From the view point of the instability of the hydroquinone compound, Kitagawa³ suggested that halenaquinone 3 might be a secondary product of halenaquinol isolation. If so, xestoquinol 2 may be a genuine natural product instead of xestoquinone 1. Recently, Schmitz⁵ isolated adociaquinone A ((+)-5), adociaquinone B ((+)-6), and related compounds from sponges of the genus Adocia in Truk Lagoon.

We and Kitagawa have previously determined the absolute stereochemistry of halenaquinone (+)-3 and halenaquinol (+)-4 as shown in Chart I by the theoretical calculation of the CD spectra of pertinent derivatives.⁶ Furthermore, we experimentally showed that the absolute stereostructures of (+)-3 and (+)-4 determined theoretically were confirmed by the total synthesis⁷ of halenaquinone (+)-3 and halenaquinol (+)-4. On the other hand, the absolute stereochemistry of xestoquinone has remained undetermined. Here, we report the first total synthesis of xestoquinone (+)-1 and xestoquinol 2 and the experimental determination of their absolute stereostructures. In addition, we also describe the theoretical confirmation of the absolute configuration of the xestoquinone series on the basis of the CD spectra of naphthalene-diene derivatives with a twisted π -electron system.

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